Spinal Cord Implantation with Acidic Fibroblast Growth Factor as a Treatment for Root Avulsion in Obstetric Brachial Plexus Palsy

Pei-Hsin Lin¹,³, Henrich Cheng¹,³, Wen-Cheng Huang¹,³, Tien-Yow Chuang²,³,*

¹Division of Nerve Repair, Department of Neurosurgery, Neurological Institute, ²Department of Physical Medicine and Rehabilitation, Taipei Veterans General Hospital, and ³National Yang-Ming University School of Medicine, Taipei, Taiwan, R.O.C.

Nerve root avulsion carries the worst prognosis among brachial plexus injuries and remains a great challenge for surgeons to repair. In this case, a boy with complete avulsion of the left-side C6 root presented with flaccid paralysis of the left arm after birth. As there was no significant spontaneous recovery, the patient underwent operation when he was 6 months old. One end of the nerve graft from the sural nerve was anastomosed with the avulsed C6 root, and the other end was implanted into the ventrolateral aspect of the spinal cord with fibrin glue containing acidic fibroblast growth factor. After 2 years of follow-up, there has been significant improvement in motor function and in electrophysiologic studies over the left upper limb. [J Chin Med Assoc 2005;68(8):392–396]

Key Words: acidic fibroblast growth factor, obstetric brachial plexus palsy, root avulsion, spinal cord implantation

Introduction

Obstetric brachial plexus palsy (OBPP) represents a special subgroup of brachial plexus injury and is one of the most devastating complications resulting from birth trauma. Complete root avulsion is a particularly serious condition with a bleak prognosis, as it is generally regarded as a type of spinal cord lesion.¹ Current surgical repair of acute injury may be accomplished by reconstruction involving nerve grafts or nerve transfer, using the surrounding nerves as donors.² The outcome, however, is far from satisfactory. As such, strategies to enhance axon and oligodendrocyte regeneration and peripheral neural regrowth are being targeted.

In our previous rat model studies, we found that several transected cervical roots could regenerate through intercostal nerve grafts using fibrin glue; adding acidic fibroblast growth factor (aFGF) could increase the efficacy of sprouting.³,⁴ We also showed that functional recovery and most regenerating axons within the dorsal horn disappeared with re-cutting of the grafted sensory roots.⁵ We therefore believe that aFGF stimulates neural proliferation and growth and may also be neuroprotective.⁶–⁸ However, the efficacy of growth factors in realistic models has not yet been supported by formal clinical studies.

Although a few studies suggested a modest benefit in return of function after avulsed roots were implanted into the ventral cord,⁹,¹⁰ there have been no publications to show the efficacy of neurotrophic factors in repairing avulsed spinal nerve roots. We present a unique case of OBPP involving nerve grafting and implantation of the completely avulsed C6 root into the ventrolateral aspect of the spinal cord using a fibrin gel containing aFGF.
Case Report

A 4-month-old patient with a birth weight of 3,170 g presented with persistent flaccid paralysis of the left upper limb after an uneventful vaginal delivery and birth history. The newborn underwent a comprehensive multidisciplinary rehabilitation program from birth, but no significant functional improvement was achieved.

A physical examination revealed mild atrophy of the left biceps, triceps and deltoid. Muscle power scored by the Medical Research Council system was graded 0/5 for over-shoulder abduction and elbow flexion, and 2/5 for elbow extension. There were no abnormalities in wrist or finger movements. Rating using the Toronto grading system by Clarke and Curtis was 0/7 (no contraction) for over-shoulder abduction and elbow flexion, 3/7 (motion > 1/2 range with gravity eliminated) for over-elbow extension with gravity eliminated, and full antigravity movement for over flexion and extension of the wrist, fingers and thumb. Grasping function was normal. Shoulder function according to Gilbert’s classification (modified Mallet scale) was stage 0 (no abduction) (Figure 1A).

Electrodiagnostic study of the left upper limb demonstrated markedly decreased motor unit recruitment for the deltoid and biceps, and moderately decreased motor unit recruitment for the triceps. Occasional fibrillation potentials were also noted over the deltid and biceps. Magnetic resonance imaging of the cervical spine revealed a pseudomeningocele on the left side at level C5–6, consistent with nerve root avulsion. No immediate surgical intervention was initiated due to parental hesitation. However, since there was no further motor improvement, the boy underwent surgery when he was 6 months old.

Under general anesthesia, the patient was placed in a prone position. An incision was made longitudinally in the midline of the posterior neck region, and was deepened to spinous processes and lamina. A C3–T1 total laminectomy revealed total avulsion of the left C6 spinal nerve root, with the distal end found extradurally (Figures 2 and 3). Intraoperative electrophysiologic study was performed during the procedure. The patient's response to sensory nerve stimulation was normal. The avulsed spinal nerve root was implanted into the spinal cord at the C4 level. Postoperative rehabilitation was initiated immediately. The patient made gradual improvement in shoulder abduction and elbow flexion. Antigravity shoulder abduction and anterior flexion to 120° was achieved 2 years after the operation (Figure 1B).
examination was performed to determine the continuity of other cervical roots.

A peripheral nerve graft, 25 mm in length, was harvested from the left sural nerve and kept in Hanks’ balanced salt solution. One end of the graft was anastomosed microscopically to the C6 nerve root with 10-O microsutures in an end-to-end fashion. The other end of the graft was then approximated into the ventrolateral aspect of the spinal cord through a tiny pia incision. Fibrin glue (Beriplast® P; Aventis Behring, Marburg, Germany) was prepared by mixing the fibrinogen (10 mg/mL) with aprotinin solution (2,000 kIU/mL) plus calcium chloride (8 mM) and aFGF (20 µg); it was applied to the surgical area to form a glue cast. After the nerve root was reconnected, fibrin glue was applied to the grafted area. To support the nerve root regenerating process and achieve significant concentrations of aFGF at the target side, we performed nerve root injections of aFGF (20 µg) through a monopolar needle at 1 and 1.5 years after the operation. This study was approved by the Medical Ethics Committee of our hospital and the local institutional review board. Informed consent was obtained from the child’s parents.

Six months after the operation, traces of shoulder abduction and elbow flexion motor activity developed. Nine months postoperatively, the boy achieved a Toronto grading of 2/7 (motion < 1/2 range with gravity eliminated) for over-shoulder abduction and elbow flexion, and 4/7 (full motion with gravity eliminated) for elbow extension. After 1 year of follow-up, the Toronto grading turned 4/7 for over-shoulder abduction and elbow flexion, and 5/7 (motion < 1/2 range against gravity) for over-elbow extension.

Electrodiagnostic study revealed increased recruitment of motor units in the deltoid, biceps and triceps, compared with previous electromyographic examination. The amplitude of the compound motor action potential in the left deltoid and biceps approximated 45% and 83%, respectively, as compared with the right-side muscles (Table 1). Two years postoperatively, over-elbow flexion was 5/7 and over-shoulder abduction and elbow extension was 6/7 (motion > 1/2 range against gravity). Shoulder function assessed by Gilbert’s classification was improved to stage IV, with shoulder abduction to 120–160° (Figure 1B). Electrodiagnostic study after 2 years of follow-up was not completed because of pain on needle examination, and the parents refused further examination.

Discussion

Although surgical intervention to regain function of avulsed roots in humans is uncommon, axons seem capable of regenerating in a favorable environment. Some earlier studies provide experimental data that point to the possibility of restoring the spinal cord and avulsed root connection. Numerous laboratory investigations in different animal species, including primates, have focused on strategies for implantation of avulsed roots into the spinal cord. Such investigations showed that reconnection between the spinal cord and injured roots can be achieved with regeneration across the peripheral and central nervous system (CNS) interface. Although activation of nerve regrowth is vital, it is only the initial element in the
complex process of regeneration. Additional components such as facilitation of axonal growth and precision of axonal regeneration must also be addressed. Whether or not neurotrophic factors are a required component of modern surgical root repair remains to be seen.

The minimal motor regain noted in this patient 6 months after surgical repair with aFGF is greater than the average rate of axonal growth observed in human nerves, suggesting that reformed functional synapses are a possible contributor to early recovery; however, collateral sprouting from neighboring nerves cannot be excluded. For ethical reasons, nerve root blocking was not performed in this boy to elucidate the speculation. Thus, at present, the origin, mechanism, or pathway underlying the recovery remains unknown. Previous research found that nerve grafts used along with exogenous aFGF improved somatosensory evoked potentials (SEPs) of cut dorsal roots. After re-transection of the repaired roots, SEPs were completely eliminated. This verified that the SEPs’ origin was from regenerated axons rather than from reinnervation of adjacent intact roots.5

Sufficient new and functional synapses and regenerating axons were required for this boy to recover some motor function. Antigravity movement developed in shoulder abduction and elbow flexion 1 year postoperatively. Two years after the operation, significant functional recovery with overhead shoulder abduction was achieved (from Gilbert’s classification stage 0 to stage IV). The fact that a full year after surgery was needed for locomotor function to return may demonstrate that the synthesis rate for new synapses, graft lengths and axoplasmic transport determine the central/peripheral nervous system synapse growth rate and the duration of motor recovery.18,19

Our patient had satisfactory, rather than full, motor recovery after 2 years; perinatal root avulsion may have led to loss of some of the axotomized motor neurons, thus limiting the possibility of full recovery. Indeed, early research indicated that the motor neuron pool depleted about 7–10% per week after spinal avulsion.20–22 Another explanation for our finding could be the short half-life of aFGF added to fibrin glue. Neurotrophic factors in general have poor pharmacokinetic profiles and short in vivo half-lives.23 Mixing aFGF with fibrin glue does allow for slow release of the growth factor; however, the access of growth factors to the CNS is largely impeded by multiple clearance processes.24 Therefore, aside from the initial local injection at the repair site, the authors performed 2 sequential C6 nerve root injections, guided by electromyographic recordings, 1 and 1.5 years after surgery. In future, advances in targeted drug delivery methods for neurotrophic factors will most likely determine the clinical usefulness of such factors for CNS conditions.

In this case, recovery may have been related to something other than reconstruction of the C6 nerve root. As we were unable to clearly demonstrate a cause and effect relationship between treatment and recovery, our case study has limitations. Animal studies provide the opportunity to use various modalities for investigations; however, experimental investigations were not possible in our patient.

In conclusion, this report indicates that nerve graft and spinal cord implantation using aFGF in fibrin glue may improve upper limb muscle activity in cervical root avulsion of OBPP, and reduce the level of disability and consequences in this patient population. However, a definitive positive effect for this novel technique requires verification in further studies involving greater numbers of patients.

Table 1. Electrophysiologic studies of the left side upper limb in a 6-month-old boy with obstetric brachial plexus palsy

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<thead>
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<th>Preoperative</th>
<th>6 months postoperatively</th>
<th>1 year postoperatively</th>
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<tr>
<td>CMAP amplitude, %*</td>
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<tr>
<td>Deltoid</td>
<td>ND</td>
<td>23</td>
<td>45</td>
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<tr>
<td>Biceps</td>
<td></td>
<td>48</td>
<td>83</td>
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<tr>
<td>Motor unit recruitment/Fib, PSW†</td>
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<tr>
<td>Infraspinatus</td>
<td>Normal/–</td>
<td>Markedly decreased/1+</td>
<td>Markedly decreased/0</td>
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<td>Deltoid</td>
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<td>Moderately decreased/0</td>
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<td>Biceps</td>
<td>Markedly decreased/2+</td>
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<tr>
<td>Triceps</td>
<td>Moderately decreased/0</td>
<td>Moderately decreased/0</td>
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<tr>
<td>ADQ</td>
<td>Normal/0</td>
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*Percent relative to normal side, with stimulation site at Erb’s point; †grading of Fib, PSW is as follows: 0, no Fib; 1+, single trains in ≥ 2 muscle regions; 2+, moderate numbers in ≥ 3 muscle areas; 3+, many in muscle regions; 4+, in all areas of sampled muscles.25 ADQ = adductor digiti quintus; CMAP = compound motor action potentials; Fib = fibrillation potentials; ND = not done; PSW = positive sharp waves.
References